
Biological Database Design

Week 3

Winter '05

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Question and Answer

- Discuss homework
- Q & A on last two weeks' material

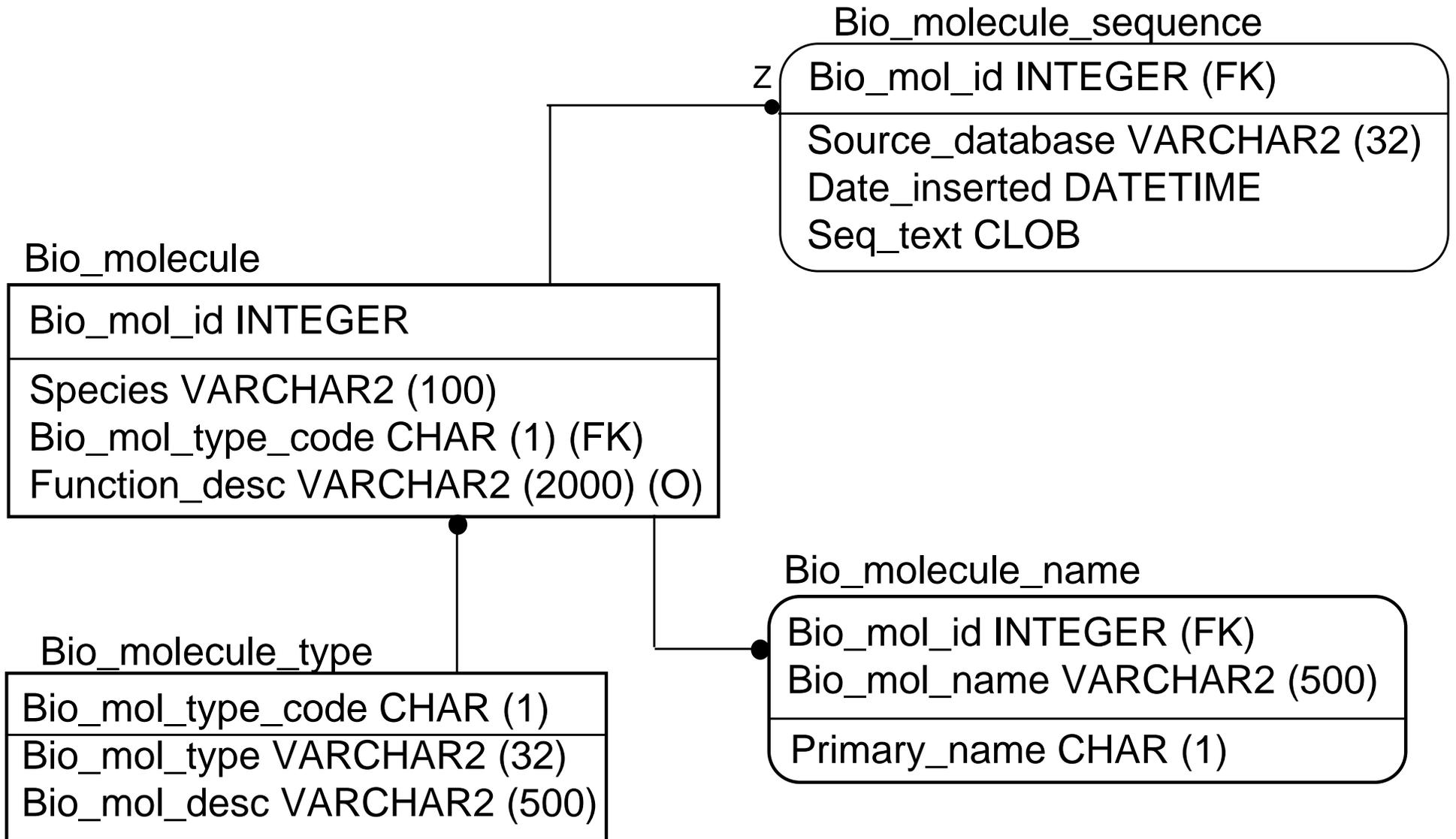
Introduction to SQL

- SQL = Structured Query Language
 - Except that the spec says SQL doesn't stand for anything
- Standard language for accessing data in relational databases
- A nonprocedural language
 - Say what you want, not how to get it
 - A RDBMS has a query optimizer that figures out how to get the data
- RDBMS purists point out that it is not fully compliant with relational database theory
 - Poor support of domains
 - Allows tables without keys

Introduction to SQL

- Data Definition Language (DDL)
 - ❑ CREATE TABLE, DROP TABLE
 - ❑ CREATE INDEX
 - ❑ Constraints: UNIQUE, PRIMARY KEY, FOREIGN KEY, NOT NULL
- Data Manipulation Language (DML)
 - ❑ INSERT, UPDATE, DELETE
 - ❑ SELECT
 - ❑ UNION, INTERSECT, EXCEPT

Example Tables



CREATE TABLE

- Use to create a table
- **CREATE TABLE** table1
(column1 datatype **PRIMARY KEY**,
column2 datatype)
- Each table should have a primary key constraint on one or more columns
- Use **UNIQUE** to enforce alternate keys

CREATE TABLE

Create a table to store biological molecules

```
CREATE TABLE Bio_molecule (  
    Bio_mol_id INTEGER PRIMARY KEY,  
    Species VARCHAR2 (50) NOT NULL,  
    Bio_mol_type_code CHAR (1) NOT NULL,  
    Function_desc VARCHAR2 (2000)  
)
```

PRIMARY KEY is equivalent to UNIQUE, NOT NULL

Other DDL Commands

■ ALTER TABLE

- ❑ Add/drop/modify a column of a table
- ❑ Not all DBMS support drop and modify

■ CREATE INDEX

- ❑ Create an index on a column or combination of columns
- ❑ Implementation detail: indexes are used by DBMS to enforce constraints and optimize lookup
- ❑ UNIQUE constraints automatically create index

■ DROP TABLE, DROP INDEX

INSERT

- Use INSERT to get data into a table
- **INSERT INTO** table1 (column list)
VALUES (value list)
- Column list is optional, but should specify it if the statement is included in application code
 - Remember, the columns in a table are not in any particular order!

INSERT

Insert the name “PTP1B” for biological molecule #1456. It is a primary name.

```
INSERT INTO Bio_molecule_name  
  (Bio_mol_id, Bio_mol_name, Primary_name)  
VALUES (1456, 'PTP1B', 'Y')
```

Text is surrounded by single quotes.

UPDATE

- Use to alter data in a table
- **UPDATE** table1
SET column1 = new value,
column2 = new value
WHERE column3 = condition
- WHERE clause is optional. Without it, the UPDATE will apply to all rows in the table

UPDATE

Change calmodulin to be the primary name.

```
UPDATE Bio_molecule_name  
SET     Primary_name = 'Y'  
WHERE   Bio_mol_name = 'calmodulin'  
AND     Bio_mol_id = 456
```

Bio_mol_id portion of where clause is probably unnecessary.

DELETE

- Removes row(s) from table
- **DELETE FROM** table1
WHERE column1 = condition
- WHERE clause is optional. Without it, DELETE will remove all rows from the table.
 - Won't remove table
 - To do this, use DROP TABLE

DELETE

Delete all Incyte sequence data

```
DELETE FROM Bio_molecule_sequence  
WHERE Source_database = 'INCYTE'
```

SELECT

- Use to get information out of tables
- **SELECT** column1, column2
FROM table1
WHERE column3 = condition
- WHERE clause is optional. Without it, the statement returns all rows in the table

SELECT

- List the primary name and bio_mol_id for all molecules:
 - ❑ SELECT Bio_mol_id, Bio_mol_name
FROM Bio_molecule_name
WHERE Primary_name = 'Y'
- List all biological molecules stored in the database:
 - ❑ SELECT *
FROM Bio_molecule

SELECT DISTINCT

- Use to get a list of distinct values
- **SELECT DISTINCT** (column1, column2)
FROM table1
- Can have one or more columns in the select statement
- Multiple columns will provide distinct combinations of values of those columns

SELECT DISTINCT

Find out what types of biological molecules are represented in the Bio_molecule table:

```
SELECT DISTINCT Bio_mol_type_code  
FROM Bio_molecule
```

JOIN

- Joins are used to combine information from multiple tables
- Two types of syntax
- **SELECT** table1.column1, table2.column2
FROM table1, table2
WHERE table1.column3 = table2.column3
- **SELECT** table1.column1, table2.column2
FROM table1
JOIN table 2 **ON** (table1.column3 = table2.column3)

JOIN

Show the biomolecule type, rather than the code, for all types represented in Bio_molecule:

```
SELECT DISTINCT Bio_mol_type
FROM Bio_molecule bm,
     Bio_molecule_type bmt
WHERE bm.Bio_mol_type_code = bmt.Bio_mol_type_code
```

```
SELECT DISTINCT Bio_mol_type
FROM Bio_molecule bm
JOIN Bio_molecule_type bmt
     ON bm.Bio_mole_type_code = bmt.Bio_mol_type_code
```

LIKE and Wildcards

- Wildcards are ‘%’ and ‘_’
 - ‘%’ = any number of characters
 - ‘_’ = exactly one character
- Used with keyword LIKE
- Select information on all biomolecules with the word “kinase” in one of their names
 - ```
SELECT bm.Bio_mol_id, Bio_mol_name, Species
FROM Bio_molecule bm,
 Bio_molecule_name bmn
WHERE bm.Bio_mol_id = bmn.Bio_mol_id
AND Bio_mol_name LIKE '%kinase%'
```

Contents of strings are case-sensitive

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# ORDER BY

- ORDER BY returns rows in order
- List the names assigned to Biomolecule #478 in alphabetical order:
  - ```
SELECT    bio_mol_name
FROM      bio_molecule_name
WHERE     bio_mol_id = 478
ORDER BY  bio_mol_name ASC
```
- ASC or DESC

Aggregate Functions

■ COUNT

- ❑ Count number of sequences from RefSeq DB
- ❑ SELECT COUNT (*)
FROM Bio_molecule_sequence
WHERE Source_database = 'RefSeq'

■ GROUP BY

- ❑ Count number of sequences from each DB
- ❑ SELECT Source_database, COUNT (*)
FROM Bio_molecule_sequence
GROUP BY Source_database

Aggregate Functions

- MAX and MIN
 - `SELECT MAX(Date_inserted)`
`FROM Bio_molecule_sequence`
 - Can be used on numeric and date fields
- SUM
- AVG

String Functions

- DBMS specific implementations
- Usually have at least:
 - Substrings
 - Length

Subqueries

- Can nest SQL statements:
 - Select all primary names for human proteins:

```
SELECT Bio_mol_name
FROM Bio_molecule_name
WHERE Bio_mol_id IN (
  SELECT Bio_mol_id
  FROM Bio_molecule
  WHERE Species = 'Homo sapiens'
  AND Bio_mol_type_code = 'P'
)
```

Subqueries

■ EXISTS

- Another way to express subsets

```
SELECT Bio_mol_name
FROM Bio_molecule_name bmn
WHERE EXISTS (
  SELECT *
  FROM Bio_molecule bm
  WHERE Species = 'Homo sapiens'
  AND Bio_mol_type_code = 'P'
  AND bm.Bio_mol_id = bmn.Bio_mol_id
)
```

Subqueries

- Can also use NOT IN and NOT EXISTS
- Choice between using JOIN, IN, or EXISTS is a performance tuning issue
- Optimizer will usually “convert” for you, but sometimes it pays to optimize, or “tune” the query yourself
- For more details:
 - SQL Performance Tuning, by P. Gulutzan and T. Pelzer

Subqueries

- Can join back to the same table
- Show the primary name for all biomolecules for which there are no other names:

```
SELECT Bio_mol_name
FROM    Bio_molecule_name bmn1
WHERE   Primary = 'Y'
AND NOT EXISTS (
    SELECT *
    FROM    Bio_molecule_name bmn2
    WHERE   Primary <> 'Y'
    AND     bmn2.Bio_mol_id = bmn1.Bio_mol_id
)
```

CLOBs

- **CLOB** = **C**haracter **L**arge **O**bject
- Implementation is very DBMS specific
- Usually do not have access to many functions
 - ❑ No substring or length functions
 - ❑ Can't use in WHERE clause
 - ❑ Can even be difficult to load in and select out

Sequence Data

- Bioinformatics has traditionally focused on handling sequence data
- Many sequence databases are not relational
 - Particularly old ones: implemented prior to good DBMS support for CLOBs
 - GenBank and Swiss-Prot: originally flat file DBs, now have some relational storage
 - Lion's SRS (Sequence Retrieval System)
 - Popular way to handle sequences
 - Flat file based

Sources of Sequence Data

- Public
 - NCBI
 - GenBank = all sequences
 - RefSeq = curated sequences
 - ExPASy
 - SWISS-PROT = highly curated protein sequences
 - TrEMBL = uncurated protein sequences (translated EMBL)
- Private
 - Incyte (out of the genomics business)
 - Celera
- Proprietary
 - In house sequencing efforts

Sequence Data

- A typical sequence “entry” contains:
 - Sequence text
 - Metadata
- Metadata is not uniform across sources
 - Will almost always have the species
 - Curated data sources will usually have
 - Meaningful name (‘Mitogen-Activated Protein Kinase’)
 - Some indication of function
 - Uncurated data sources are often annotated by computer
 - Names often “similar to protein X” or “hypothetical protein”

Molecule to Sequence Relationship

- The same “protein” or “gene” can be represented by multiple sequence entries
- Different databases often have slightly different sequences
 - Start codon selection
 - Initiator methionine included or not
 - SNPs (single nucleotide polymorphisms)
 - Sequencing errors
 - Splice variants (a headache in their own right)

Molecule to Sequence Relationship

- Difficult to ascertain when two sequences are the “same” molecule
- Requires scientists to set appropriate rules for your database
 - I’ve used 90 – 95% identity over at least 50 residues
 - Exact cutoffs depend on need for accuracy vs. need for inclusiveness
- Some databases bypass the issue and treat each sequence individually
 - Potential for lots of data duplication
 - Decision is ultimately made based on database scope

Relational Implementation

Bio_molecule

Bio_mol_id INTEGER
Bio_mol_type_code CHAR(1) (FK)
Species_id INTEGER (FK)

Bio_sequence

Bio_sequence_id INTEGER
Bio_mol_id INTEGER (FK)
Source_id INTEGER (FK)
Source_identifier VARCHAR2(50)
Date_inserted DATETIME
Sequence_text CLOB

Sequence_source

Source_id INTEGER
Source_name VARCHAR2 (100)
Source_desc VARCHAR2 (500)
Source_url VARCHAR2 (500) (O)

Sequence Text

- Protein and nucleotide
 - Nucleotides translate to proteins at 3 base pairs per amino acid
 - DNA sequences contain introns: unexpressed DNA “inserted” into gene
- Large range in size of sequence text
 - Common to study ESTs (~300 – 500 base pairs)
 - Smallest proteins are ~50-200 amino acids
 - Largest protein is titin, which has ~27,000 amino acids
 - Genomic DNA can be millions of base pairs long

Searches on Sequence Text

- Exact match
 - ❑ Not very useful, because small variations can occur in sequences that are scientifically “the same”
 - ❑ Used to remove (or flag) obvious redundancies
 - ❑ Some uses in intellectual property
- Global match (e.g., ClustalW)
 - ❑ Finds optimal alignment over entire length of two sequences
 - ❑ Allows insertions and substitutions
 - ❑ Not good at identifying matching regions within sequences that also have unmatched regions

Searches on Sequence Text

- Local match (e.g., BLAST)
 - Most common method of searching sequence DBs
 - Looks for regions of alignment within two sequences
 - Allows insertions and substitutions
- Motif or domain searches
 - Look for regions of sequence that match known patterns
 - Used to infer function
 - Search for characteristic motifs (BLOCKS, PRINTS, PROSITE)
 - Search for domains (Pfam, SMART)
 - Allow insertions and substitutions

Sequence Searching in RDBs

- Can't perform searches on CLOBs
- No easy way to implement the most useful types of searches in standard SQL
- Not all substitutions are equal
 - Some substitutions are more “conservative” than others
 - Preserve basic chemical properties of amino acid
 - Use a “substitution matrix such as BLOSSUM to specify “cost” of substitutions
 - Choice of substitution matrix may depend on personal preference, goals of project

Sequence Searching in RDBs

- Usually search on sequence text outside of relational database
- BLAST runs on a “database” of sequences in FASTA format
- Two options
 - Store sequences in database, but dump to FASTA for BLAST
 - Store sequences in FASTA flat files, reference these in database
 - Either way, DB and flat files can get out of sync
 - Storing sequences in database makes DB “gold standard”
- Oracle 10g implements BLAST searches in the database

Sequences as Non-Atomic Data

- In some databases, sequences are split into a table in which each amino acid or base pair is a row
- This is done when there is a need to store data about individual positions in the sequence
- Intermediate solutions: “break out” certain regions to store as individual residues
 - Functional motifs
 - Duplicates data

Sequence Metadata

- Metadata = data about data
 - Sequence is primary data
- Some metadata is a property of a particular sequence
 - Biophysical measurements: isoelectric point, extinction coefficients
- Some metadata is a property of the gene or protein that the sequence represents
 - Biological data: function, subcellular localization
- Species metadata can go either way
 - Depends on how you choose to handle orthologs in your database
 - Messiness of functional variation among orthologs means that a protein/gene is usually best associated with a single species

Sequence Species

- Species data is really a hierarchy
- For most applications, storing the full hierarchy is out of scope
 - Exceptions
 - Evolutionary biology
 - If need ability to perform deep searches on species (for “all mammals”, etc.)
- Usually need at least scientific name and one common name
 - Some people will also provide basic classifications: specifics depend on scope of DB
- Can link to/incorporate NCBI’s taxonomy DB
 - www.ncbi.nlm.nih.gov/Taxonomy

Sequence Function

- Two types of function (at least!)
 - Biochemical
 - The chemical process for which the protein/gene is responsible
 - Examples: kinase, calcium-binding
 - Enzymes: cross-reference EC (Enzyme commission) numbers (ENZYME: <http://www.expasy.org/enzyme/>)
 - Non-enzymes and enzymes: cross-reference molecular function Gene Ontology (<http://www.geneontology.org>)
 - Cellular/Process
 - The cellular pathway or process in which the protein/gene participates
 - Examples: DNA repair, long term potentiation
 - Cross-reference biological process Gene Ontology

Sequence Function

- Link to disease states may be considered a type of function, too
 - ICD codes (<http://www.who.int/classifications/icd/en/>)
- One gene or protein may be involved in multiple biochemical and cellular functions
 - Many enzymes have multiple binding sites
 - Many signal transduction proteins participate in multiple pathways
- There are always exceptions to standard ontologies
- If a scientist's favorite gene doesn't fit the standard ontology, and he can't explain why, he won't store the data!
 - Always provide a comment field

Additional Metadata

- Too numerous to list
 - Chromosome
 - Ligand binding sites
 - Intron locations
 - Active site residues
- Highly dependent on interests of group using database
- Often difficult to classify
- Constantly expanding list
- Some text, some numeric

Metadata Issues

- Due to incomplete nature of biological research, the features that are available vary widely by molecule
 - If you try to make a table with a column for each feature, you will have a lot of NULLs
 - Alternatively, making each feature its own table leads to an explosion of tables in your schema

Additional Metadata

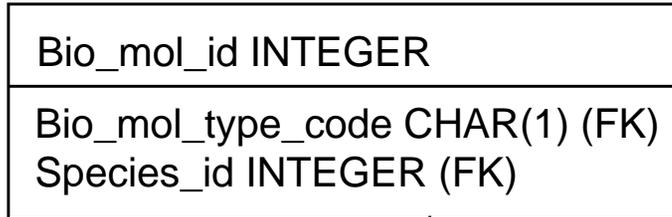
- Most public databases handle additional metadata as “feature table”
 - GenBank/EMBL feature table
 - Each feature has a location (optional: without location, feature is assumed to apply to entire sequence)
 - Features have “keys” (identifying names)
 - Features can have qualifiers (in GenBank spec, some are mandatory)
 - Example: primer-binding site feature
 - Key = primer_bind
 - Optional qualifiers: allele, citation, db_xref, evidence, gene, label, locus_tag, map, note, standard_name, PCR_conditions
 - Swiss-Prot has similar feature design
 - Comments apply to entire sequence
 - Examples: function, tissue specificity
 - Features are assigned a location
 - Examples: domain, binding site, post-translationally modified residue

Entity-Attribute-Value Design

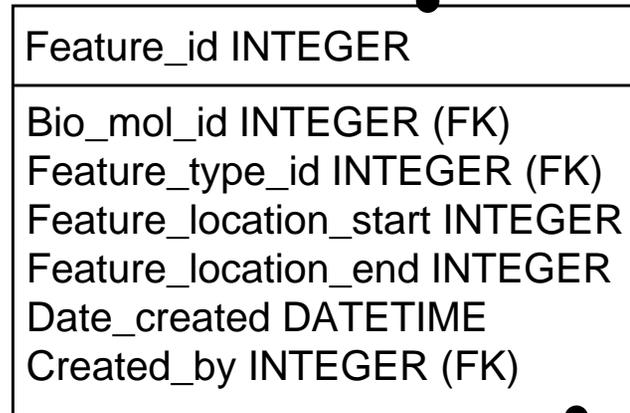
- Standard design pattern used in many fields
- Values in table specify the feature, feature qualifier, and feature value
- If database needs to store features that apply only to regions of the sequence, add a “location” column
 - Requires separate tables for feature and qualifier, to avoid duplicating location
- Consider making feature type and feature qualifier lookup tables
 - Prevents duplicate names for same feature
- Store text and numeric features separately
 - Preserve ability to use numeric aggregate functions
 - Store units of numeric features

Relational Implementation

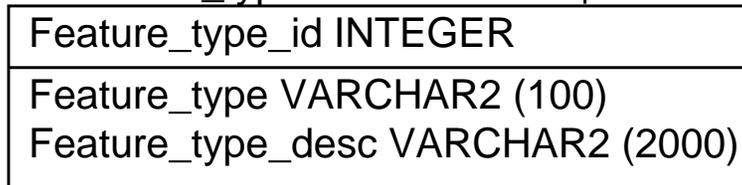
Bio_molecule



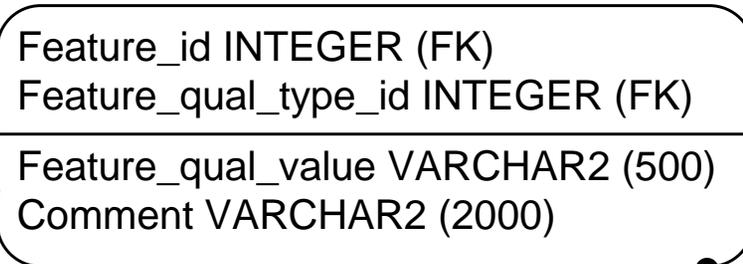
Feature



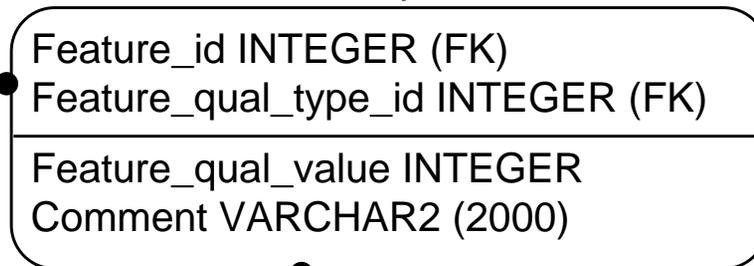
Feature_type



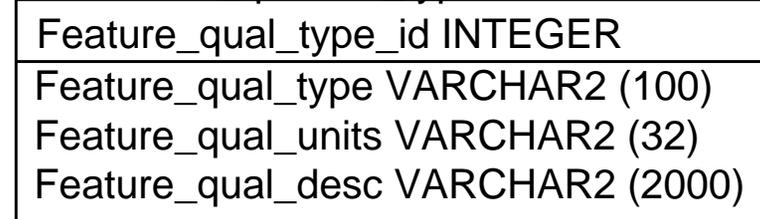
Text_feature_qualifier



Numeric_feature_qualifier



Feature qualifier type



Difficulty Classifying Biological Data

- Biology is often a very “fuzzy” science
- Data is incomplete: scientists are constantly forming and discarding hypotheses
- Nature has a seemingly infinite way of combining features
- Dilemma
 - “Fuzziness” is real and important
 - Need “hard” classifications to support truly deep queries
 - Compromise
 - Make classification system user-extensible
 - Provide comment fields into which all of the real ambiguity can be entered

Tracking the Source of Data

- It is often desirable to track the source of features
 - Particularly if features may be entered by users (rather than downloaded from source databases only)
 - Also desirable because different source databases may provide contradictory metadata
- Lack of “feature source” tracking has created a problem with function annotations in public databases
 - Sequence A is annotated as a kinase because of sequence similarity with Sequence B
 - Sequence B turns out not to be a kinase
 - More likely: Sequence A has same basic structure as Sequence B, but lacks kinase function
 - Sequence C is annotated as a kinase because of similarity to Sequence A
 - If none of the “function transfers” are traceable, the function annotations cannot be trusted

Tracking the Source of Data

- In science, it is important to be able to lookup and evaluate source reference
- Science is incomplete
 - Your research contradicts the data in the database
 - Which is in error? Are both right, and we don't see the full picture yet?
 - Scientist needs to return to original source and evaluate the experiment

Tracking the Source of Data

- Gold standard is publication in peer reviewed journal
- Usually, but not always, indexed in PubMed (www.ncbi.nlm.nih.gov/PubMed)
- Other sources
 - Chemistry journals
 - Dissertations (rarely read, let alone cited...)
 - Webpages
 - Internal company reports

Tracking the Source of Data

- Reference data is actually quite complex
 - In many applications, it is enough to link to PubMed
 - I usually provide ability to create internal, non-structured reference object for things not indexed in PubMed
 - If need to allow queries into references, must store the reference itself
 - Find all features supported by papers on which Joe Q. Scientist is an author
 - NCBI allows downloading of an XML version of reference, which is easy to parse into your database
 - Object Management Group Bibliographic Query Service (OMG-BQS) model
 - <http://industry.ebi.ac.uk/openBQS/>
 - class diagram is in the specification section
-

Sequence Versioning

- Some public databases now version their sequences
 - Example: RefSeq
 - Sequence is identified by an accession number and a version
 - NM_005842.2
 - In general, only latest version of sequence is available
- Must decide how to handle versioning in your database
 - Keep all versions or latest version only?
 - If you keep all versions, do you associate different versions of the same sequence with each other?
 - What happens to any metadata added to the sequence when a new version comes out?

Questions to Ask

- Is your primary interest the sequences or the proteins/genes they represent? (Or both?)
 - Tells you whether you can simplify one or the other
- Do you need to search over “aggregate” species designations?
 - Tells you how much of the species hierarchy you need to store
- Do you need to search on details of supporting data, or just link to it?
 - Tells you whether you need to store all reference data, or just a link to it
- Do you need to associate data with a particular version of a sequence?
 - Tells you whether you need to track versions

Additional Data Models

- ENSEMBL data model
 - Relational database for ENSEMBL
 - http://www.ensembl.org/Docs/schema_description.html
- bioSQL
 - <http://obda.open-bio.org>
 - From the Open Bioinformatics Foundation (open-bio.org)
- aMAZE
 - Interesting data model for representing function
 - <http://www.amaze.ulb.ac.be>
 - Representing and analysing molecular and cellular function using the computer. J. van Helden, et. al. (2000) Biol. Chem. 381:921-935.

Homework

- Reading for this week's class
 - GenBank portion of the NCBI handbook, UniProt user manual (on website)
- Homework: Project plans are due next week

- Reading for next week's class
 - Paper discussing GeneLogic's approach to managing gene expression data
 - Implementing LIMS: A "How To" Guide
- Optional reading for next week's class
 - Nature Genetics paper on MIAME (strongly recommended, but will require a trip to the library)
 - A computer scientist's explanation of microarrays (strongly recommended for those not familiar with the technique)
 - MAGE-ML paper